

Cell sorting using spectral flow cytometry

The Emory Pediatrics/Winship Flow Cytometry Core recently acquired a new BD cell sorter, replacing the FACS Aria II. The new BD S8 imaging + cell sorter and the Cytex Aurora CS cell sorter are spectral, therefore we suggest the following strategies to ensure successful sorts:

Optimize cell concentration



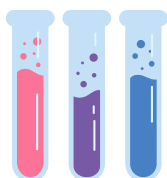
Final cell preparation should be as concentrated as possible, up to 10-20 million cells per mL. If you have less than 2 million then re-suspend in 300uL of liquid. Live cells should be kept on ice as much as possible from the moment they are isolated until they are run on the machine. Once stained, cells should be kept in the dark.

Filter the sample



Filtering the sample before sorting is essential to eliminate clumps and debris that could clog the sorter. Use a cell strainer [PN 352235, Falcon] or a nylon mesh with a pore size of no more than 50 μm to filter the sample.

Bring appropriate controls



Unstained cells: An unstained cell control is required for determining cellular autofluorescence. In this way, unmixing algorithms can isolate and extract the spectral contribution of autofluorescence from the rest of the spectral signatures and improve signal resolution.

Single stain controls: Both cells and beads can be used and should be tested to determine which control provides the most accurate fluorophore spectral signature for unmixing.

Use appropriate buffer solutions



Final buffers should include BSA or FBS, DNase solution to help prevent aggregation, and HEPES buffer [[Flow Cytometry Protocols To Prevent Sample Clumping](#)]. For cells that are generally "sticky" and more likely to clump together, a small amount of EDTA can be added to help abate this problem.

Feel free to contact the core with any additional questions or for consultation purposes: ajrae@emory.edu

[Emory Pediatrics/Winship Flow Cytometry Core web page](#)

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